

PROJECT REPORT

On

**SYNTHESIS OF QUERCETIN BASED SCHIFF BASE AND ITS
ANTI-INFLAMMATORY ACTIVITIES**

Submitted by

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*In partial fulfillment for the award of the
Post graduate Degree in Chemistry*



**DEPARTMENT OF CHEMISTRY
AND
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**DEPARTMENT OF CHEMISTRY
AND
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ST. TERESA'S COLLEGE (AUTONOMOUS)
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This is to certify that the project "SYNTHESIS OF QUERCETIN BASED SCHIFF BASE
AND ITS ANTI-INFLAMMATORY ACTIVITIES" is the work done by
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**DEPARTMENT OF CHEMISTRY
AND
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CERTIFICATE

This is to certify that the project work titled “**SYNTHESIS OF QUERCETIN BASED SCHIFF BASE AND ITS ANTI-INFLAMMATORY ACTIVITIES**” is the work done by **JAYALAKSHMI J.** and **SONA MARY MARTIN** under the guidance of **Dr. Shanty A. A. Assistant Professor** Department of Chemistry and Centre for Research, St. Teresa's College, Ernakulam in partial fulfilment of the award of the Degree of Master of Science in Chemistry at St. Teresa's College, Ernakulam affiliated to Mahatma Gandhi University, Kottayam.

Project Guide

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Project Guide

DECLARATION

I hereby declare that the project work entitled **“SYNTHESIS OF QUERCETIN BASED SCHIFF BASE AND ITS ANTI-INFLAMMATORY ACTIVITIES”** submitted to Department of Chemistry and Centre for Research, St. Teresa's College (Autonomous) affiliated to Mahatma Gandhi University, Kottayam, Kerala is a record of an original work done by me under the guidance of **Dr. Shanty A. A. Assistant Professor,** Department of Chemistry and Centre for Research, St. Teresa's College (Autonomous), Ernakulam. This project work is submitted in the partial fulfillment of the requirements for the award of the Degree of Master of Science in Chemistry.

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Chapter 1

Introduction

1.1 SCHIFFBASE

The discovery of new chemotherapeutics with novel bioactivities and functionalities to fight current emerging diseases has become the most significant research in pharmaceutical science. Schiff bases are adaptable pharmacophores that can chelate with metals in a variety of oxidation states to produce complexes. The discovery of new chemotherapeutics with novel bioactivities and functionalities to fight current emerging diseases has become the most significant research in pharmaceutical science.

Schiff bases are the complexes synthesized from the condensation of primary amines with carbonyl groups which are used in various fields (Figure 1.1). Hugo Schiff originally wrote about them in 1864. Schiff bases are versatile C=N (imine) containing compounds having a broad spectrum of biological activities. And they also showed some degree of antibacterial, anti-inflammatory activities. Mainly Schiff bases are used as intermediates for the synthesis of amino acids and also used as ligands for the preparation of metal complexes [1].



Figure 1.1: General Scheme of formation of Schiff Base

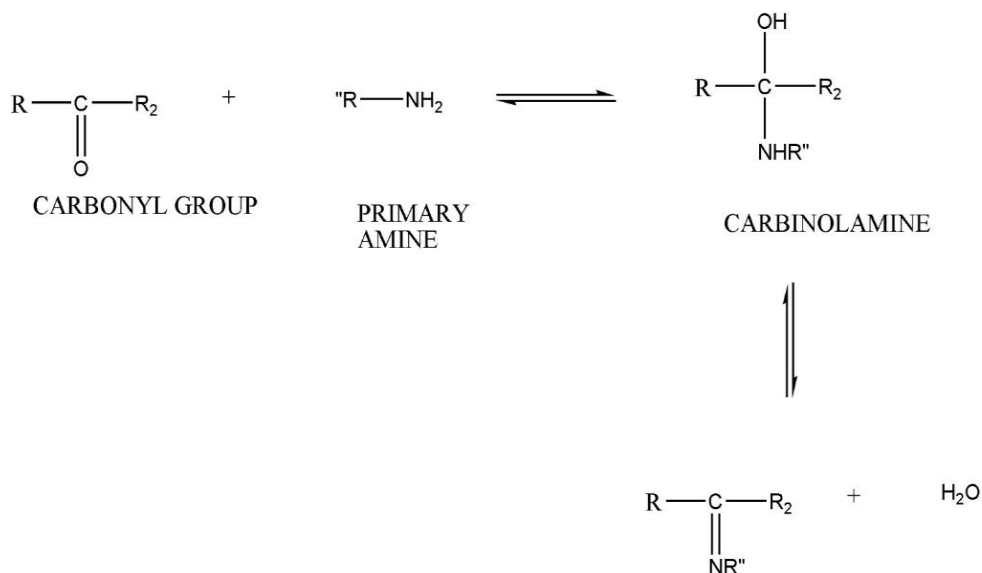


Figure1.2: Mechanisms showing the formation of Schiff Base

Aldehyde or Ketone combines with primary amine to give carbinolamine where the double bond between C and O gets broken and leads to the formation of an OH bond. The alcoholic group Carbinolamine which is an unstable addition compound undergoes acid or base catalyzed dehydration. Following the removal of water, Carbinolamine gives an Imine with C=N bond (Figure1.2). The reaction of forming Imine is a reversible reaction. Addition followed by elimination are the two types of reaction that take place in the formation of Schiff Base [2].

The azomethine group, which is shared by Schiff Base compounds, has the generic formula $\text{RHC}=\text{NR}_1$, where R and R₁ are heterocyclic, cycloalkyl, or aryl groups that can be substituted in many ways. These substances are also known as imines, or azomethines. Aryl substituted Schiff bases are more stable compared with alkyl substituted because of its alternative double bonds. Research has shown that the presence of a single pair of

electrons in the nitrogen atom's SP^2 hybridized orbital of the azomethine group has important chemical and biological ramifications. Because of their special C=N group trait, synthetic flexibility, and relative ease of synthesis, Schiff bases are often excellent chelating agents. The versatility of Schiff base ligands and the uses of their complexes in biology, chemistry, and industry make more research in this field extremely desirable [3].

The most Important research in pharmaceutical science today is the development of innovative chemotherapeutics with unique bioactivities and functionality to combat newly emerging diseases. Because of their numerous and valuable scientific applications, Schiff base metal complexes have been the focus of coordination chemistry research during the course of several decades of rigorous study on metal-based pharmaceuticals. They may be used as antibacterial, antimicrobial, anticancer, antiviral, anti-inflammatory, analgesic, antifungal, and many other medicinal medicines.

Medicinal resistance has become a global problem in medical research in recent years due to the majority of pathogenic organisms being able to deactivate medicinal compounds. For this reason, it Requires urgent attention from chemical and pharmaceutical scientists to address these severe challenges of multidrug resistance [4].

1.2 BIOLOGICAL APPLICATIONS OF SCHIFF BASE

A carbonyl group and an amino group condense to form Schiff Base ligands and their complexes. The resulting metal complexes may possess antioxidant, antifungal, and antibacterial qualities [5]. In many enzymatic processes where an enzyme interacts with an amino or carbonyl group of

the substrate, Schiff Base appears to be a crucial intermediary. In organic chemistry, it has several synthetic applications as well. Acylation of Schiff Bases by acid anhydride, acid chlorides and acyl cyanides is initiated by the attack on the nitrogen atom and it leads to the net addition of the acylation agent to the CN double bond. These exhibit substantial antibacterial action as well. The metal complexes of the Schiff Bases have much better antibacterial activity than their free ligands [6]. For improving the product yield Schiff Base metal complexes can be used which have strong catalyst activity. For the peroxidation of several alkenes including cyclohexene, cis and trans stilbene, cyclooctene, the complexes of V, Mn, Fe, Co, Cu and Zn ions were utilized as catalysts [7].

1.2.1 ANTIDEPRESSANT ACTIVITY

Schiff bases of isonicotinoyl hydrazone N-[(1Z)-substituted aromatic)methylidene] pyridine-4-carbohydrazides were found to have considerable antidepressant and nootropic properties in vitro. Compounds substituted with nitro, halogen, and dimethoxy groups also had strong antidepressant properties [8].

1.2.2 ANTIMALARIAL ACTIVITY

The malaria genus is caused by Plasmodium. In general, Plasmodium is made up of four species. P. Vivax, P. Falciparum, P. Ovale, and P. Malaria. Serious health issues could result from malaria. Nowadays, the hunt for novel medications to treat this illness is essential. Antimalarial medications can be made from Schiff bases. N-[(1E)-(5-nitro-1-naphthyl)methylene]-1-(2-(tri-fluoromethyl)phenyl)methanamine was the

most effective antimalarial agent among 5 nitroisoquinoline Schiff bases[9].

1.2.3 ANTIINFLAMMATORYACTIVITY

One of the body's main defense mechanisms against infection, poisonous substances, allergies, and other unpleasant stimuli is inflammation[9]. In many chronic conditions, an unchecked and ongoing inflammation may be a contributing cause [4]. Additionally, medicines with Anti Inflammatory properties are imidazole groups that contain Schiff base transition metal complexes.

Conversely, diseases caused by microorganisms, such as bacteria or fungi, are typically benign and increase gradually in fatality [10]. NSAIDs and analgesics, which are often used Anti Inflammatory medicines, have been linked to a number of side effects, including myocardial infarction [11], congestive heart failure, nausea and vomiting, dyspepsia, stomach ulceration/bleeding, diarrhea [12], hypertension, and retention of salt and fluids. Numerous biological effects, including those that are anti-inflammatory, analgesic, antiviral, antipyretic, antirheumatic, and antibacterial, have been demonstrated by the pyrazolone derivative 4-aminoantipyrine (4-amino-1,5-dimethyl-2-phenylpyrazole-3-one) and its derivatives. Mohammad Sayad Alam, Jung-Hyun Choi, Dong-Ung Lee conducted experiments on this derivative. These groups of substances also function as potent inhibitors of the synthesis of prostanoid, platelet thromboxane, and cyclooxygenase isoenzymes [13].

1.2.4 ANTIOXIDANTACTIVITY

Antioxidants are naturally occurring chemicals that protect living organisms from damage caused by harmful molecules called free radicals. These are produced by the cells in the body in response to free radicals [14]. Free radicals play an important role in the pathogenesis of many diseases, including cancer, diabetes, liver damage, autoimmune diseases, heart disease, atherosclerosis, and aging [15]. Therefore, antioxidants with the potential to scavenge free radicals play an important role in the treatment and prevention of these diseases [16]. Antioxidants are often used as catalysts for antibiotics. Antioxidant compounds have a high ability to scavenge free radicals. Antioxidants play an important role in retarding or preventing the oxidation of oxidizable substances (substrates) [17], [18]. In vivo, antioxidant compounds prevent damage to 4,444 macromolecules and cells by interfering with free radical molecules [19]. Therefore, the importance of searching for antioxidants has increased dramatically in recent years [20]. Currently, synthetic antioxidants are widely used as they are cheap and effective compared to natural antioxidants [21].

1.2.5 ANTICANCERACTIVITY

By condensing salicylaldehyde with 2-amino-4-phenyl-5-methyl thiazole, a Schiff base ligand was created. In good yield, the ligand forms compounds with Zn(II), Cu(II), Ni(II), and Co(II). Elements analysis, magnetic susceptibility, molar conductance, infrared spectra, ¹H and ¹³C NMR, mass, electronic absorption, and ESR spectroscopy were used to describe the produced compounds. In order to compare the anticancer activity of the synthesized compounds with that of doxorubicin as a

reference medication, the compounds' anticancer activity was investigated against many human tumor cell lines, including breast cancer MCF-7, liver cancer HepG2, lung carcinoma A549, and colorectal cancer HCT116. In comparison to the inhibition in the four cell lines (HepG2, MCF7, A549, and HCT116), the study demonstrated that Zn(II) complex exhibited potent inhibition against human TRK by the ratios of 80, 70, 61, and 64%, respectively [22].

1.3 IMPORTANCE OF REAGENTS USED

1.3.1 QUERCETIN

Family of secondary plant chemicals known as polyphenols is composed of rings of phenolic compounds. Polyphenols are known to be present in plant-based diets. They are typically found in plant-based products like tea, coffee, red wine, and chocolate in addition to a variety of fruits and vegetables. Recent research has demonstrated the preventive role of plant polyphenols against cancer, cardiovascular disease, and neurological illnesses. They are strong antioxidants that guard against oxidative damage. These polyphenols are secondary metabolites of plants and are essential parts of defense mechanisms in plants [23].

"Quercetin" is the Latin name for the flavonoid, and it means "oak forest." The human body does not manufacture this kind of flavonol. The benefits of quercetin for ailments like arthritis, cancer, allergic responses, inflammation, and cardiovascular disease have long been established. Moreover, mitochondrial biogenesis, platelet aggregation, and lipid peroxidation all depend on flavonoids [20]. A vast array of seeds, nuts, flowers, bark, and leaves can be found in nature. It is found in plants that are related to tomatoes, canola, berries, onions, leeks, grapes, apples, and

tea [24]. In addition to ginkgo biloba, hypericum perforatum, and elderberries, the primary sources of quercetin are onions, apples, and tea [24]. The formula for it is $C_{15}H_{10}O_7$.

The polar auxintransport inhibitor is a naturallyoccurring substance.The oxygen atomon the first carbon ofthe quercetin molecule is basic and can react with strong acids to form salts. The molecule also contains a ketocarbonyl group.Its molecular structure consists of four active groups: an o-dihydroxy group (B), an inter-ring dihydroxy group (A),adouble bond(C2),aC3ring,anda4-carbonylgroup.Quercetinhas strong antioxidant qualities since it has phenolic hydroxyl groups and double bonds. Furthermore,studies carried out in vitro and in vivo have shown quercetin's antibacterial activity in addition to its effective suppression of related gene expression, anticancer activity, antiangiogenic activity, and other processes that aid in the formation of biofilms.Additionally, quercetin plays a major role in loweringmycotoxins, which protect cells from damage.

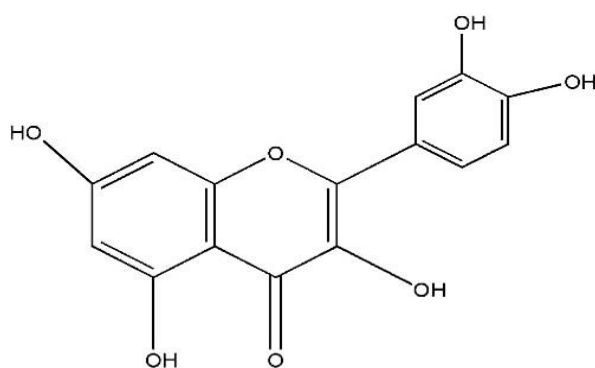


Figure1.3:StructureofQuercetin

Quercetin is pentahydroxyflavone having five hydroxy groups. Three cyclic rings present in the structure among them two of them are phenyl groups. 2 hydroxy group present in each phenyl rings. The centre ring have a double bond at its second and third position, fourth position contains carbonyl group and there also a hydroxy group present in the third carbon. Thus it have a molecular formula of $C_{15}H_{10}O_7$ (Figure 1.3). The presence of Phenolic hydroxyl groups and double bonds endows quercetin with a strong antioxidant activity.

1.3.2 PARABROMOANILINE

p-Bromoaniline has a molecular formula C_6H_6BrN (Figure 1.4). It is a chemical in which the para position of an aniline molecule is replaced with a bromine atom. This substance is readily accessible for purchase and can be utilized as a building block, for example, in the Gomberg-Bachmann process to prepare p-Bromobiphenyl. 4-Dihydroquinazolines and azo dyes are made using bromoaniline as a precursor. It is further added to the *Moraxella* sp. strain G growth medium as a carbon and nitrogen supplement [25].

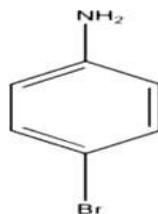


Figure 1.4: Structure of p-Bromoaniline

1.3.3 2-AMINO-4-NITROPHENOL

2-amino-4-nitrophenol has a molecular formula $C_6H_6N_2O_3$ (Figure 1.5). 2,4-dinitrophenol reacts with sodium sulfide, 2-amino-4-nitrophenol is created (Farris, 1978). Agfa produced it for the first time in 1898 (Society of Dyers and Colourists, 1971). At the moment, hair coloring products use about 150 kg of 2-amino-4-nitrophenol.

The study examined the fungicidal properties of 2-amino-4-nitrophenol and its derivatives, which are produced by substituting various chemical radicals for the hydrogen atom in its amino group. When an aldehyde group replaces the hydrogen atom in an amino group, the fungicidal activity against *Rhizoctonia solani* and *Bipolaris sorokiniana* increases. A ketone group substituted for the hydrogen atom amplifies the inhibitory impact on *Venturia inaequalis* and *Sclerotinia sclerotiorum* [26].

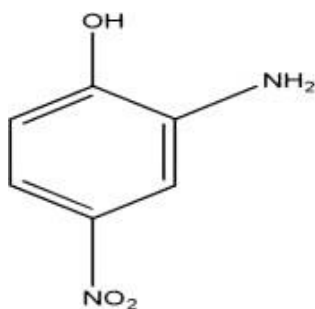


Figure 1.5: structure of 2-Amino-4-nitrophenol

1.3.4 2-AMINO-4-METHYLPHENOL

2-amino-4-methylphenol has a molecular formula C_7H_9NO (Figure 1.6). It remains a sensitizer in contact with allergens. Disperse Yellow 3 is 2-amino-4-methylphenol. In 100% ethanol, it combines with acetylacetone to produce 4-(2-hydroxy-5-methylphenyl)imino-2-pentanone. It was changed by refined human hemoglobin into dihydrophenoxazinone.

Tridentate with biological activity Schiff base ligands have been synthesized and studied using IR, elemental analysis, electronic spectrum data, molar conductance, and 1H NMR. These ligands are synthesized by reacting isatin with 2-amino-4-methylphenol and their metal complexes, Ti (IV), Zr (IV), Cd (II), and Hg (II). Using disc diffusion techniques, the antibacterial activity of the Schiff base and its metal complexes has been investigated [13].

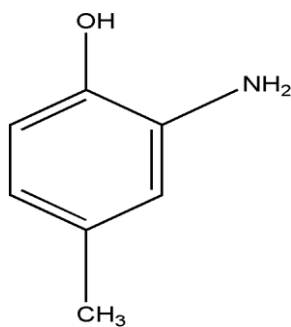


Figure 1.6: Structure of 2-Amino-4-methylphenol

1.3.5 ORTHOPHENYLENEDIAMINE

The formula for the chemical compound O-phenylenediamine is $C_6H_4(NH_2)_2$ (Figure 1.7). An essential precursor to numerous heterocyclic compounds is this aromatic diamine. OPD is a white substance, however

airborne oxidation causes samples to seem darker. When combined with m- and p-phenylenediamine, it is isomeric.

Carbon nanodots (C dots) were created using a straightforward hydrothermal process using o-phenylenediamine (OPD). Carbon nanodots made from o-phenylenediamine are employed in cells to sense Cu^{2+} ions. When stimulated at 420 nm, the C dots show photoluminescence at 567 nm. Cu(OPD)_2 complexes occur on the surfaces of C dots when Cu^{2+} ions are present, causing the color of the dots to change from yellow to orange and increasing the PL intensity [27].

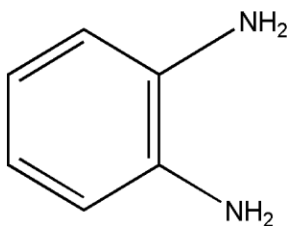


Figure 1.7: Structure of o-Phenylenediamine

1.4 ANTI-INFLAMMATORY STUDIES OF SYNTHETIC SCHIFF BASE

1.4.1 INFLAMMATION

Inflammation is the response of our body's immune system against the foreign body that enters into our body. Whenever our body encounters an outside invader like a bacteria or virus, toxin or our body suffers an injury, our immune system gets activated and releases the substances into the blood called inflammatory markers. The job of these inflammatory markers is to trap the virus or bacteria or heal the injured tissue in case of an injury. These responses may be in the form of redness or swelling of the wound.

There are two different types of Inflammation. ACUTE and CHRONIC inflammation. Acute inflammation is usually short term. Symptoms of acute inflammation include pain, redness, tenderness or swelling at the site of the infection or injury. Actually, acute inflammation is a protective response of the body that helps in healing the body from an infection or injury.

When the body keeps triggering the immune system to release white blood cells and other inflammatory markers even when there are no outside dangers, this inflammation is called chronic inflammation. Chronic inflammation is usually long lasting and it can go from a few months to a few years. The symptoms of chronic inflammation are little vague and might include abdominal pain or discomfort, joint pain, fever etc, which may cause dangers to our body [28].

1.4.2 CAUSES OF INFLAMMATION

Acute inflammation can result in:

- Pain or soreness;
- Blushed skin where the damage occurred.
- Swelling.
- Temperature.

Inflammation that is chronic can last for months or even years. It is associated with or may be associated with a number of disorders, including:

Diabetes, cardiovascular disease (CVD), arthritis and other joint diseases, allergies and chronic obstructive pulmonary disease (COPD), rheumatoid arthritis, psoriasis [29].

1.4.3 PREVENTION OF INFLAMMATION

The first and most effective Anti-Inflammatory measure that can follow is good food, nutrition, proper exercise etc. The top Anti-Inflammatory food is turmeric. The compound curcumin in turmeric has been studied a lot for combating inflammation in the body. Other compounds with effective Anti-Inflammatory properties are lutein in green leafy vegetables, omega-3 fatty acids in fatty fish and walnuts, the green mung bean also has been found to have excellent Anti-Inflammatory properties. Foods that should be completely avoided to prevent blood inflammation or reduce existing inflammation are processed foods, trans fat (Vanaspati Dalda), avoid alcohol etc. There is no perfect test for detecting inflammation. Usually doctors and medical professionals recommend a test when inflammation is associated with other medical conditions. Doctors treat inflammation by

putting patients on nonsteroidal Anti Inflammatory drugs like Ibuprofen or aspirin [30].

1.4.4 ANTI-INFLAMMATORY DRUGS AND ITSEFFECTS

Non-steroidal anti-inflammatory drugs (NSAIDs), which act via inhibition of the cyclooxygenase (COX) isozymes, were discovered more than 100 years ago. They continue to be an essential part of the pharmacological treatment of both acute and persistent pain. The biological activities of the COX-1 and COX-2 isozymes are distinct; analgesic effectiveness is predominantly (but not completely) linked to COX-2 inhibition, whereas the inhibition has distinct adverse effects. All available NSAIDs, including acetaminophen and aspirin, are associated with potential side effects, particularly gastrointestinal and cardiovascular effects, related to their relative selectivity for COX-1 and COX-2 [31].

Ibuprofen was first made available to people as an anti-inflammatory medication in England in 1967 and the US in 1974. Milligram for milligram, it possesses strong but mild anti-inflammatory qualities comparable to aspirin, but with much less detrimental effects on the stomach. Although ibuprofen, fenoprofen, and naproxen have a chemical relationship, this does not imply that none of the other members of this class of propionic-acid derivatives have any impact. The medication's analgesic qualities are most likely a result of its anti-inflammatory qualities. It is a nonsteroidal agent since it prevents the manufacture of prostaglandins and does not affect the adreno pituitary axis. Ibuprofen has been shown to be effective in rheumatoid arthritis and osteoarthritis and is probably effective in ankylosing spondylitis, gout, and Bartter's syndrome [32].

Chapter 2

Literature Review

Amandeep Sodhi and his co-workers conducted a study on systemic autoimmune illness, rheumatoid arthritis (RA) and the effect of quercetin alone or mix with methotrexate on inflammation. The temporomandibular joint (TMJ) may be impacted by RA, despite the fact that the hands, wrists, and feet are the joints most commonly affected. Disease-modifying antirheumatic drugs (DMARDs) have antiproliferative, immunosuppressive, and anti-inflammatory activity, controlling the progression of RA. At the moment, methotrexate (MX) is the initial course of treatment for RA. The continuous MX dosing may induce pertinent adverse effects, including liver and renal damage, leukopenia, and drug resistance. Evidence suggests QT is a treatment for RA that can be used because it increases synovial cell apoptosis in cell culture, suppresses pro-inflammatory mediators, and modifies preclinical studies have shown that activating transcription factor signaling pathways improves clinical symptoms and lowers pro-inflammatory cytokines in RA patients' plasma levels. The arthritic animal treated with QT and/or MX showed a significant increase in the nociceptive threshold when compared to untreated arthritic animals. Results suggest that QT reduces inflammation and shows hepatoprotective effect in an experimental model of RA in TMJ [33].

Four common flavonoid glycoside compounds—luteolin, kaempferol, apigenin, and quercetin—are present in many plants with a

variety of biological activity. Ruxia Wang, Yu Chang, Xin Liu, and some coworkers analysed these 4 flavonol glycoside compounds and measured the NO level, phagocytosis, DPPH and ABTS radical scavenging activities, and ferric reducing antioxidant power. The current study concentrated on their anti-inflammatory and antioxidant activities *in vitro*. This investigation showed that all four drugs could lower phagocytosis and NO concentrations. Their antioxidant activities increased as the concentration increased. The IC₅₀ ABTS values were 0.59, 0.8506, 0.8243, 0.5083, 1.4497 and 2.1563 µg/ml for luteolin, kaempferol, apigenin, quercetin, BHT and VC, respectively. They concluded that quercetin is an ideal antioxidant and anti-inflammatory drug that may be used as an adjuvant treatment for oxidative stress and inflammatory illnesses, according to test results. Furthermore, this study provided preliminary evidence that the number of phenolic hydroxyl groups directly correlates with antioxidant activity. Furthermore, compounds containing enol groups demonstrated higher antioxidant and anti-inflammatory activity compared to those lacking enol group [34].

Marija Lesjak, Ivana Beara and their colleagues analysed six quercetin derivatives. Six quercetin derivatives (quercetin-3-O-glucuronide, tamarixetin, isorhamnetin, isorhamnetin-3-O-glucoside, quercetin-3,4'-di-O-glucoside, quercetin-3,5,7,3',4'-pentamethyl ether) were evaluated for their antioxidant and anti-inflammatory properties in comparison to the activity of standard onion extract, which is the primary source of dietary quercetin, and benchmarks (butylated hydroxytoluene and aspirin). The quercetin compounds exhibited noteworthy bioactivities that were comparable to those of onions and standards. The antioxidant efficacy of quercetin was reduced upon derivatization of its hydroxyl groups.

Nevertheless, there was no clear relationship between quercetin's ability to

prevent the synthesis of inflammatory mediators and the quantity of free hydroxyl groups in it. In summary, the systemic circulation of quercetin derivatives following quercetin consumption may function as strong antioxidants and anti-inflammatory agents, as well as enhance the overall biological activity of a diet rich in quercetin [35].

Using a mouse model of asthma, A. P. Rogerio, L. H. Faccioli and their colleagues examined the anti-inflammatory properties of quercetin and isoquercitrin. After receiving two intranasal ovalbumin challenges, BALB/c mice received an immunization (ovalbumin/aluminum hydroxide, s.c.). After the initial immunization, the mice were given daily gavage of either quercetin (10 mg/kg) or isoquercitrin (15 mg/kg). Positive control: 1 mg/kg, s.c. of dexamethasone was given. Twenty-four hours following the final ovalbumin challenge, leucocytes were examined in lung parenchyma, blood, and bronchoalveolar lavage fluid (BALF). We examined interleukin-5 (IL-5) in lung homogenates and BALF. Effective eosinophilic inflammatory suppressors, quercetin and isoquercitrin may have therapeutic value in the treatment of allergies [36].

Potential application of the antioxidant quercetin to shield cells from intracellular Ca^{2+} overload: a model experiment conducted by Yumiko Nishimura and his coworkers. It is well known that quercetin shields cells from oxidative damage. One of the processes that causes cell death is the elevation of intracellular Ca^{2+} content brought on by oxidative stress. Thus, they postulated that quercetin might shield cells that are experiencing an excess of intracellular calcium ions. Using rat thymocytes and a flow cytometer fitted with appropriate fluorescence probes, the effects of quercetin on cells experiencing intracellular Ca^{2+} excess and oxidative stress were investigated in order to test the theory. The concentrations of quercetin (1–30 μM) used to protect cells from

intracellular Ca^{2+} overload caused by a calcium ionophore were comparable to those used to protect cells from H_2O_2 -induced oxidative stress. When external Ca^{2+} was removed, the cell death caused by H_2O_2 and calcium ionophore, respectively, was greatly reduced. Moreover, quercetin dramatically postponed the Ca^{2+} -dependent cell death process while having no discernible effect on the increase in intracellular Ca^{2+} concentration caused by calcium ionophore and H_2O_2 , respectively. Conclusion is that, Despite an increase in intracellular Ca^{2+} concentration, quercetin can shield cells from oxidative damage. The findings imply that quercetin is also employed to shield cells from intracellular Ca^{2+} excess [37].

Shankar A N and his co-workers examined the *in vitro* antibacterial activity of quercetin and its 1-6 derivatives against *Bacillus cereus*, *Salmonella* spp., *Escherichia coli*, and *Staphylococcus aureus* in addition to its *in vitro* antioxidant activity against DPPH free radical. With the exception of compounds 3 and 4, which had increased antibacterial activity against *Escherichia coli* when compared to quercetin, the results showed that all the investigated derivatives of quercetin had less antioxidant and antibacterial activity than quercetin itself. They came to the conclusion that quercetin had better antioxidant and antibacterial activity than its 16 derivatives at the tested concentrations. However, the *in vitro* antibacterial activity results for compounds 3 and 4 show a significant improvement in the gram-negative antibacterial activity against *Escherichia coli* when compared to quercetin [38].

Ali Dehghani and his associates published a study in the *Journal of Molecular Liquids* in 2020, volume 309, page 113035. The protective behavior of quercetin, a flavonoid group of polyphenols, was thoroughly

investigated in the study using tests and precise modeling (including molecular/atomic-level simulations, MD/MC augmented with DFT) in HCl acidic environment (containing 1 M). The Quercetin compound's structural characterisation was achieved by the application of both UV-Vis and FT-IR techniques. FE-SEM, AFM, and contact angle analyses were used to investigate the surface protective features. Results showed that steel is protected from corrosion attacks by the coating and shielding provided by quercetin molecules. After one hour of metal submission, protection degrees of roughly 95% and 93%, respectively, were attained based on the EIS and weight loss measures. Through a combined cathodic/anodic mechanism, quercetin molecules can restrict corrosion activities, as demonstrated by the potentiodynamic polarization curves. Furthermore, it was observed that the formation of a mono-protective layer was guaranteed by the adsorption of quercetin molecules in accordance with the Langmuir isotherm. The DFT simulations suggest that a donor-acceptor interfacial mechanism may be involved in the interactions between the Quercetin/iron complexations and the target metallic adsorbent. Furthermore, the MC/MD techniques guaranteed the adsorption of metal-organic complexes on the iron surface [39].

The work done by Moamen S Refat and his coworkers aimed to assess the anti-inflammatory and antioxidant properties of the novel $[\text{Ru}(\text{Q})(\text{Cl})_2(\text{H}_2\text{O})_2]$ complex (Ru(III)/Q). A novel important combination comprising ruthenium (III) ions and the flavonoid molecule quercetin (Q) was created. Investigation of the Ru (III)/Q complex using infrared (FTIR) spectroscopy revealed that Q is coordinated as a bidentate with Ru metal ions. The Ru(III) complex's octahedral geometry was identified by the magnetic susceptibility value (1.85 B.M.) and electronic (UV-Vis) spectra. According to the thermogravimetric research

(TG/DTG), the Ru(III)/Q compound is fairly stable up to 300 °C. 60 male rats were divided into six groups in order to measure biological activity. The comet assay was carried out in the brain tissue, while cytokines were measured in the testicular and brain tissues using histological and ultrastructural investigations. Administration of Ru(III)/Q, either by alone or in conjunction with DG, inhibited apoptotic activities and brought oxidative damage down to normal levels. Ru(III)/Q thus decreased oxidative stress in male rats and prevented damage to the brain and testes. D-galactose (DG) induced aging neurotoxicity, reproductive toxicity, and antihepatic cancer action are all significantly mitigated by the Ru(III)/Q complex [40].

It is commonly recognized that the genus *Bifidobacterium* has positive health effects. Yoshiaki Yokada, Yoshikazu Tsuzuki and their coworkers found that *Bifidobacterium adolescentis* secreted more anti-inflammatory compounds when exposed to quercetin and related polyphenols. Their study looked into the properties of the anti-inflammatory compounds that *B. adolescentis* secretes. Quercetin-infused *B. adolescentis* culture supernatant lowered activated macrophage levels of inflammatory mediators. While the elevation of anti-inflammatory activity by quercetin persisted after quercetin washout, spontaneous quercetin degradation was unable to promote anti-inflammatory activity. The culture supernatant's bioactive components may be heat-stable, non-phenolic, acidic biomolecules with molecular weights less than 3 kDa, according to physicochemical analysis. There is no impact of acetate and lactate on the generation of nitric oxide. When combined, *B. adolescentis*'s anti-inflammatory compounds might be tiny molecules rather than short chain fatty acids. These results led to the provisional identification of stearic acid as a bioactive candidate chemical [41].

Chapter 3

Materials and Methods

3.1 REAGENTS

All the reagents were used as analytical grade, purchased from Sigma Aldrich and were used as received.

- i. Quercetin
- ii. Parabromoaniline
- iii. 2-amino-4-nitrophenol
- iv. Orthophenylenediamine
- v. 2-Amino-4-methylphenol

3.2 SOLVENTS

Solvent used for the synthesis and purification procedure were purchased from spectrochem Ltd and used without further purification.

- i. Methanol
- ii. Ethanol
- iii. Petroleum Ether
- iv. DMSO

3.3 PHYSICAL-CHEMICAL METHODS

3.3.1 ELEMENTAL ANALYSIS

CHN Elemental Analysis provides a measure of carbon, hydrogen, and nitrogen elemental content in a sample. It can be used on a wider range of

sample types including solids, liquids, and volatile substances. It helps to determine the structure of the sample substance by knowing the composition of the elements. It gives a pure and accurate measurement of the elements present in a sample. CHN elemental analysis is based on the combustion of the sample. Upon combustion, the sample generates uniform compound gases of elements C, H, and N. Then it is measured using gas chromatography [42].

CHN Elemental Analysis was recorded at the SAIF, Cochin University of Science and Technology, Kochi, India.

3.3.2 INFRARED SPECTROSCOPY

IR is the electromagnetic radiation with a longer wavelength than visible light. IR Spectroscopy studies the interaction between matter and IR radiation. Two compounds will never show similar IR spectra. Compound or sample have change in dipole moment during vibration is the major requirement of IR Spectroscopy. Since each type of link has a unique natural frequency of vibration, not two molecules with different structures will have exactly the same infrared absorption pattern, or infrared spectrum, because two molecules of the same type in two distinct compounds are in two slightly different environments [43].

An infrared spectrometer, or more accurately a spectrophotometer, is the device that measures a compound's absorption spectrum. The organic laboratory often uses two types of infrared spectrometers: dispersive and Fourier transform (FT) devices. These two kinds of equipment offer compound spectra in the shared range of 4000 to 400 cm^{-1} . For a given molecule, the two generate virtually similar spectra; however FT provides the spectrum more rapidly than dispersive instruments.

Infrared spectra were recorded in the range 4000-400 cm^{-1} at Department of Chemistry, Bharata Mata College, Thrikkakara, Kochi, India.

3.3.3 ULTRAVIOLET SPECTROSCOPY

In the sections of the electromagnetic spectrum known as the ultraviolet (UV) and visible (VIS) regions, or the range of wavelengths from 190 nm to 800 nm, the majority of organic molecules and functional groups are transparent. As a result, absorption spectroscopy is not very useful in this wavelength range. Nevertheless, these parts of the spectrum can occasionally yield useful information. When paired with the specifics offered by nuclear magnetic resonance (NMR) and infrared spectra, that data can reveal important structural proposals. The main principle is the Beer Lamberts Law. Molecules containing both bonding and non-bonding electrons absorb energy and get excited to higher energy levels. There can be four possible types of transitions ($\pi\text{-}\pi^*$, $n\text{-}\pi^*$, $\sigma\text{-}\sigma^*$, and $n\text{-}\sigma^*$), and the following is their order of energy: $\sigma\text{-}\sigma^* > n\text{-}\sigma^* > \pi\text{-}\pi^* > n\text{-}\pi^*$ [44].

Ultraviolet-Visible Spectroscopy was recorded in the range 200 to 700 nm at Department of zoology, St. Teresa's College (Autonomous), Ernakulam, Kochi, India.

3.3.4 NUCLEAR MAGNETIC RESONANCE SPECTROSCOPY

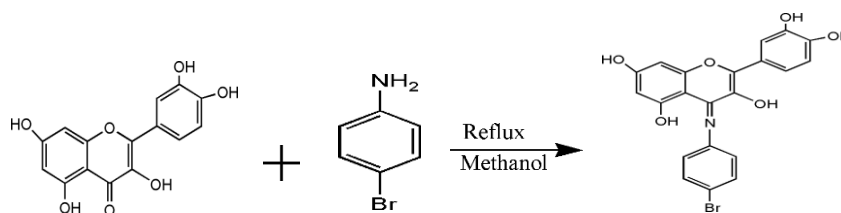
Nuclear Magnetic Resonance spectroscopy is a technique used to study molecules. It involves recording the interactions between radio frequency electromagnetic radiation and the nuclei of molecules in a strong magnetic field. NMR Spectroscopy is used in research and quality control. It can determine the purity and content of a sample as well as its molecular structures. It also analyses mixtures that contain known compounds. NMR

Spectroscopy is based on the reorientation of atomic nuclei with non-zero nuclear spin in an external magnetic field. NMR Spectroscopy provides both quantitative and qualitative data on the composition of a sample [45]. NMR spectroscopy was recorded in the range 0-20 ppm for ^1H NMR and 0-220 ppm for ^{13}C NMR at Department of Applied Chemistry, Cochin University of Science and Technology, Kalamassery, Kochi, India.

3.4 SYNTHESIS OF SCHIFF BASE

3.4.1 Synthesis of Schiff Base from Quercetin and p-Bromoaniline

Quercetin in methanol and p-Bromoaniline in methanol were combined in a 1:1 ratio and heated for 6 hours under reflux in a RB flask. (Scheme 1). After cooling and being concentrated, the resultant solution was given time for slow evaporation. After filtering, collecting, and washing with ethanol, the precipitate that had a light green hue was recrystallized and dried. The crystals were collected (Figure 3.1).



Scheme 1: Preparation of QPBA

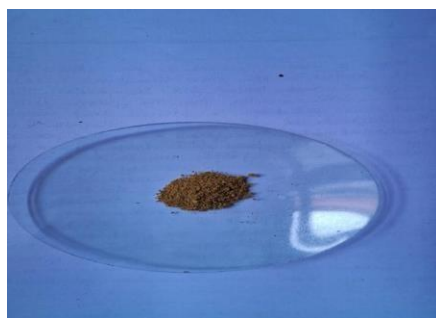
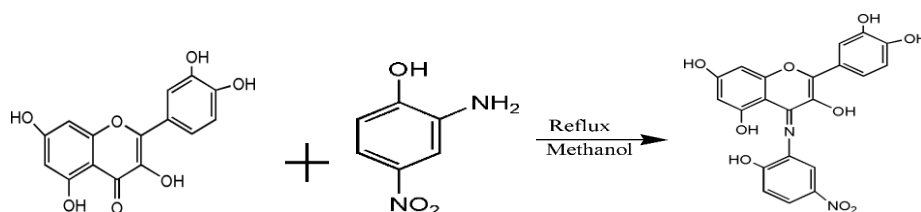


Figure 3.1: Crystals of QPBA

3.3.2 Synthesis of Schiff Base from Quercetin and 2-amino-4-nitrophenol

Quercetin in methanol and 2-Amino-4-nitrophenol in methanol were combined in a 1:1 ratio and heated for 6 hours under reflux in a RB flask. (Scheme 2). After cooling and being concentrated, the resultant solution was given time for slow evaporation. After filtering, collecting, and washing with ethanol, the precipitate that had a greenish yellow hue was recrystallized and dried. The crystals were collected (Figure 3.2).



Scheme 2: Preparation of QANP

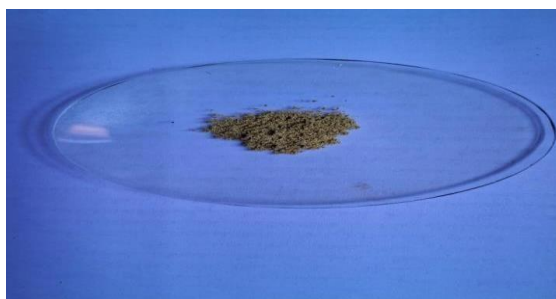
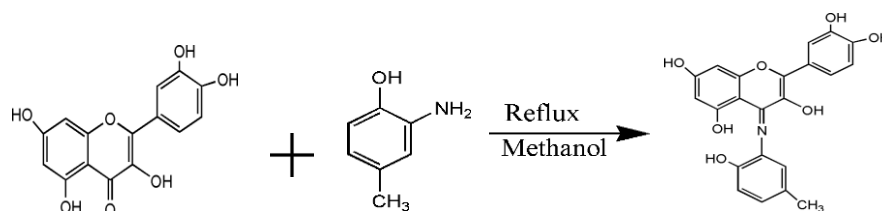


Figure 3.2: Crystals of QANP

3.3.3 Synthesis of Schiff Base from Quercetin and 2-Amino-4-methylphenol

Quercetin in methanol and 2-Amino-4-methylphenol in methanol were combined in a 1:1 ratio and heated for 6 hours under reflux in a round-bottom flask (Scheme 3). After cooling and being concentrated, the resultant solution was given time for slow evaporation. After filtering, collecting, and washing with ethanol, the precipitate that had a brownish yellow hue was recrystallized and dried. The crystals were collected (Figure 3.3).



Scheme 3: Preparation of QAMP

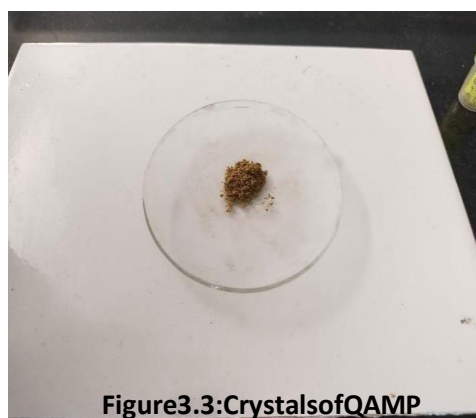
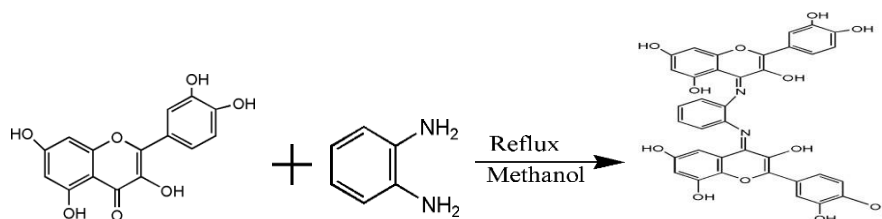


Figure 3.3: Crystals of QAMP

3.3.4 Synthesis of Schiff Base from Quercetin and o-Phenylenediamine

Quercetin in methanol and o-Phenylenediamine in methanol were combined in a 1:1 ratio and heated for 6 hours under reflux in an RB flask (Scheme 4). After cooling and being concentrated, the resultant solution was given time for slow evaporation. After filtering, collecting, and washing with ethanol, the precipitate that had a light brown hue was recrystallized and dried. The crystals were collected (Figure 3.4).



Scheme 4: Preparation of QOPD

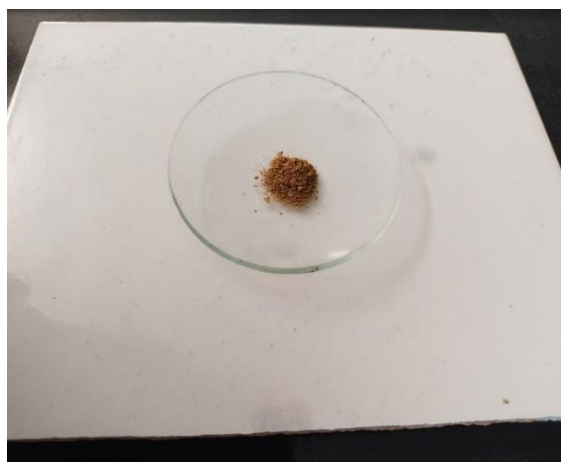


Figure 3.4: Crystals of QOPD

3.4 ANTIINFLAMMATORY STUDY OF SCHIFF BASE

3.4.1 MATERIALS USED

- i. Human Blood
- ii. Alsever Solution

- iii. Isosaline
- iv. Schiffbases

3.4.2 PROCEDURE FOR ANTI-INFLAMMATORY ACTIVITY

Plant extracts' ability to reduce inflammation was evaluated using the in vitro HRBC membrane stabilization technique. Fresh whole human blood (10ml) was collected and transferred to the heparin zed centrifuged tubes. The collected blood was mixed with an equal volume of Alsever solution (dextrose 2%, sodium citrate ,0.8%, citric acid 0.05%, sodium chloride 0.42%, and distilled water 100mL) and centrifuged with isosaline (0.85%, dissolve 8.5g NaCl in water). Autoclave 15 min at 121°C. Cool to room temperature. To 1mL of HRBC suspension, an equal volume of plant extracts in three different concentrations (10 mg/ml, 5 mg/ml and 2.5 mg/ml) was added. All the assay mixtures were incubated at 37°C for 30 Minutes and centrifuged. Using a spectrophotometer set at 560 nm the amount of hemoglobin in the supernatant solution was calculated. The percentage of protection can be hence calculated from the equation as given,

$$\text{Percent of protection} = 100 - \frac{\text{OD of test}}{\text{OD of control}} \times 100$$

Chapter 4

Results and discussion

4.1 ELEMENTAL ANALYSIS

The Elemental Analysis obtained matches with the assigned chemical formula of the proposed structure of Schiff Bases. The calculated C, H, N percentage of the Schiff Bases QPBA, QANP, QOPD and QAMP are given in (Table 4.1):

Compound	Empirical Formula	Formula weight	Colour	Observed and Calculated (%)		
				C	H	N
QPBA	$C_{21}H_{14}BrNO_6$	456.24	Light Green	55.28 (55.31)	3.09 (3.12)	3.07 (3.1)
QANP	$C_{21}H_{14}N_2O_9$	438.34	Greenish Yellow	57.54 (57.57)	3.22 (3.25)	6.39 (6.42)
QOPD	$C_{36}H_{24}N_2O_{12}$	676.58	Light Brown	63.91 (63.94)	3.58 (3.61)	4.14 (4.17)
QAMP	$C_{22}H_{17}NO_7$	407.37	Brownish Yellow	64.86 (64.89)	4.21 (4.24)	3.44 (3.47)

Table 4.1: Elemental Analysis

4.2 INFRARED SPECTROSCOPY

The peaks shown in the IR Absorption spectrum give important information about the different functional groups present in the Schiff Base (Figure 4.1 – 4.4). The peak obtained in the range $1690\text{--}1640\text{ cm}^{-1}$ [46], indicates the presence of the imine ($\text{C}=\text{N}$) group. The peaks obtained in the $3600\text{--}3200\text{ cm}^{-1}$ [47], range indicate the phenolic (OH) group. FT-IR spectral bands of Schiff bases and their spectra are given in (Table 4.2).

Compound	$\nu(\text{OH})$	$\nu(\text{C}=\text{N})$
QPBA	3286.45	1662.95
QANP	3588.05	1661.96
QOPD	3385.77	1662.95
QAMP	3372.18	1661.35

Table 4.2: Infrared spectral data of Schiff Bases

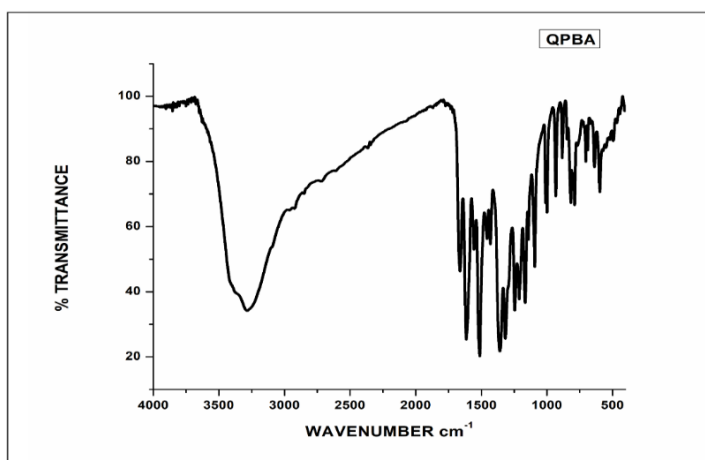


Figure 4.1: IR spectrum of QPBA

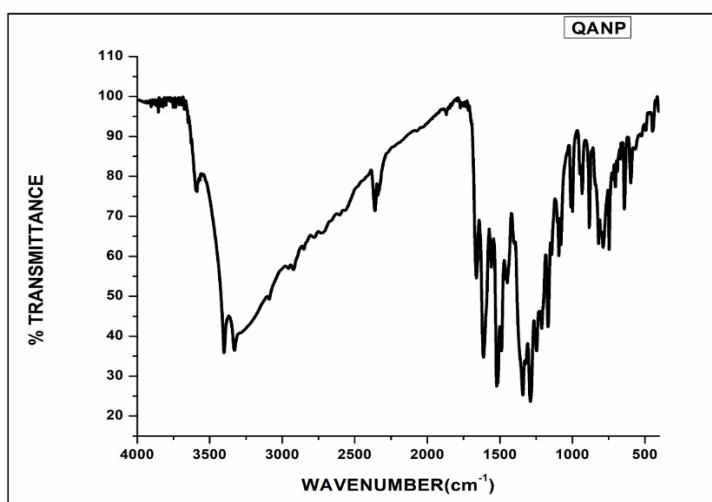


Figure 4.2: IR spectrum of QANP

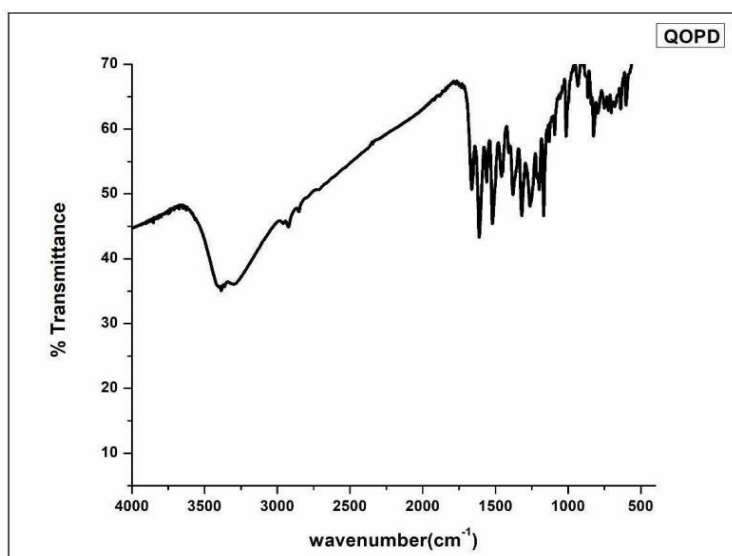


Figure4.3:IR spectrum of QOPD

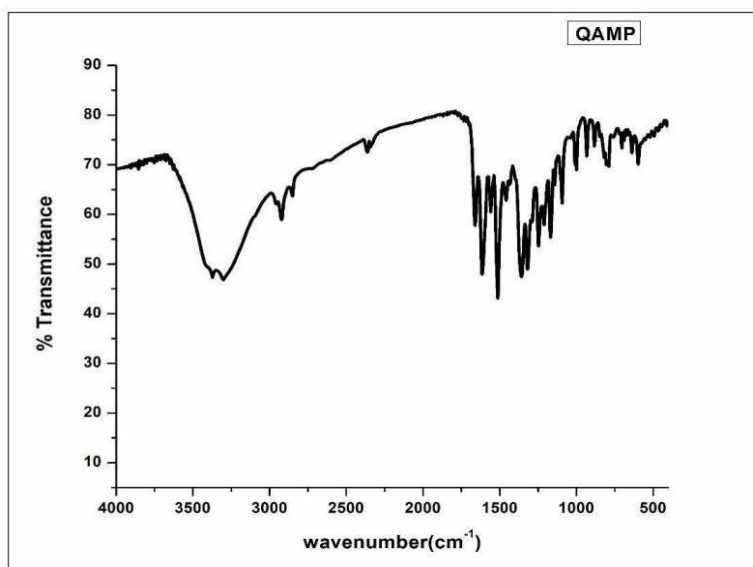


Figure4.4:IR spectrum of QAMP

4.3 UV-VISIBLE SPECTROSCOPY

UV-Visible Spectroscopy is used to study the electronic structure and its dynamics in atom and molecules. The UV-Visible spectra of compounds QPBA, QANP, QOPD and QAMP were taken in Methanol (Figure 4.5-4.8). The range 200-260 nm is due to $\pi - \pi^*$ transitions [48]. And 350-400 nm represent $n - \pi^*$ transitions [49]. The spectral data are given in the (Table 4.3):

Compound	$n - \pi^*$	$\pi - \pi^*$
QPBA	369	254
QANP	372	256
QOPD	372	256
QAMP	369	256

Table 4.3: UV visible spectral data of Schiff Bases

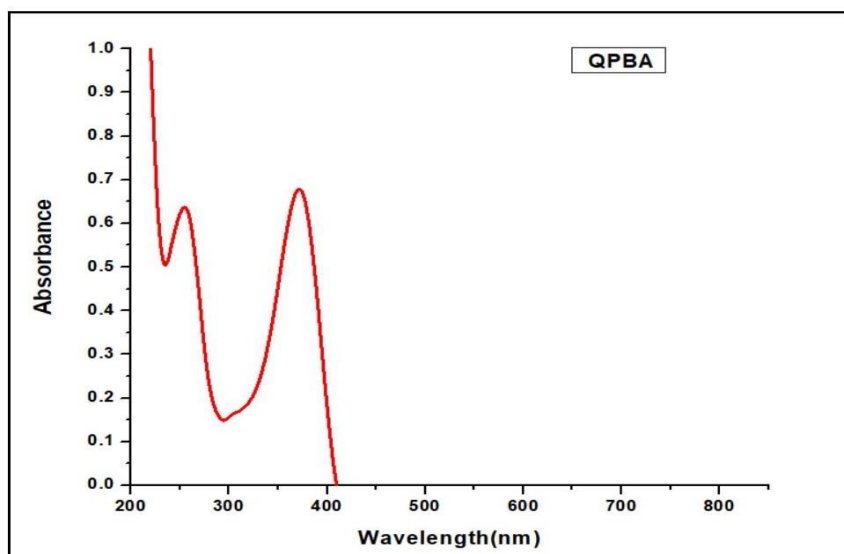


Figure4.5:ElectronicspectrumofQPBA

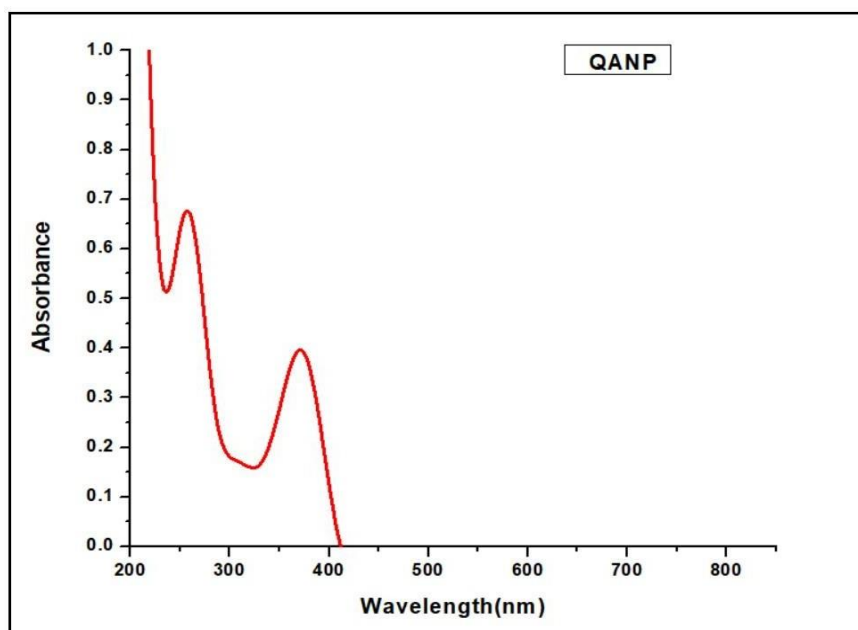


Figure4.6:ElectronicspectrumofQANP

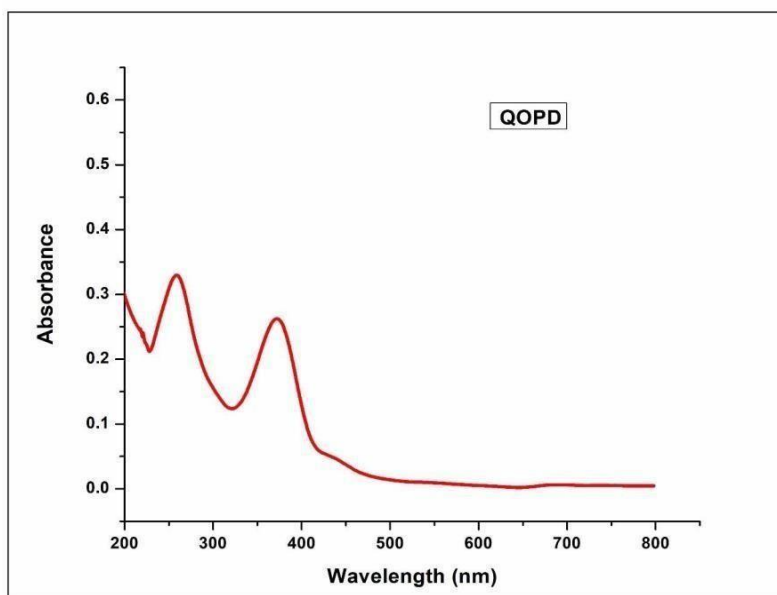


Figure 4.7: Electronic spectrum of QOPD

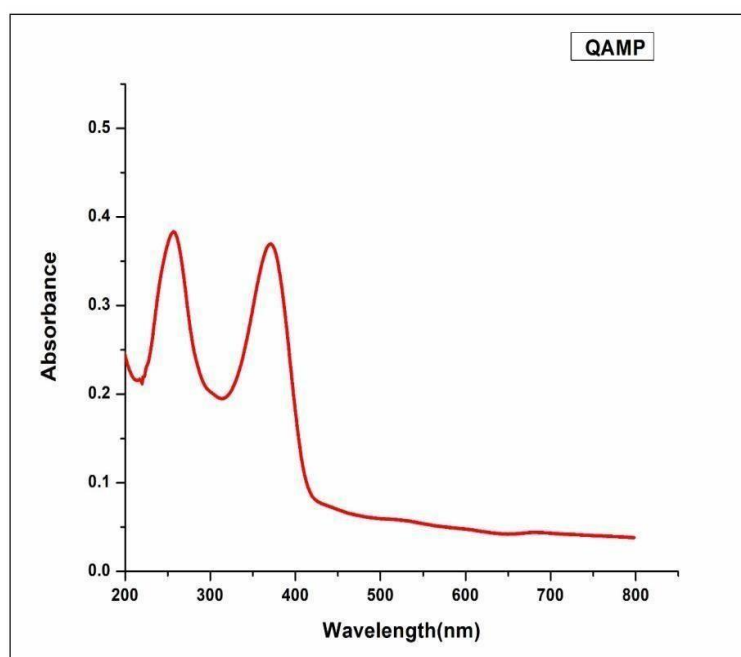


Figure 4.8: Electronic spectrum of QAMP

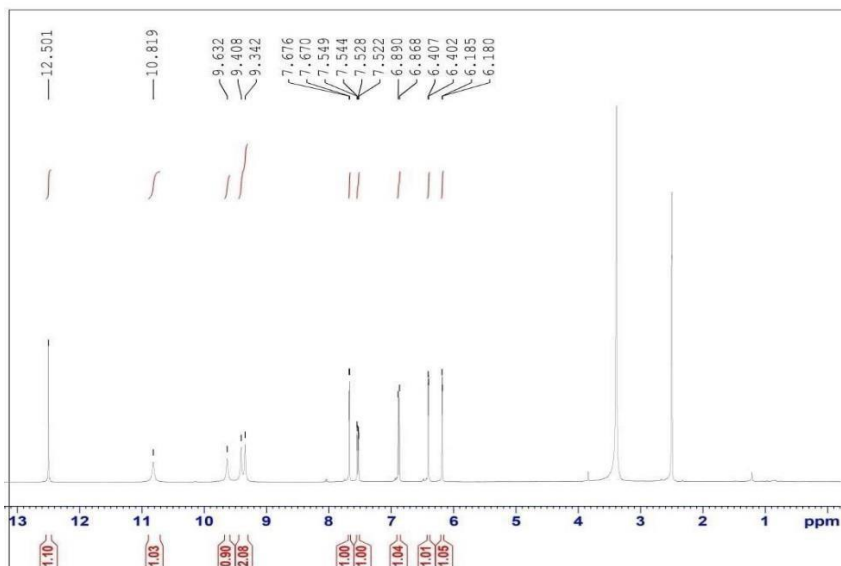
4.4 NUCLEAR MAGNETIC RESONANCE SPECTROSCOPY

NMR can be used to determine molecular conformation in solution as well as study physical properties at the molecular level such as conformational exchange, phase changes, solubility, and diffusion. The Schiff base's ^1H and ^{13}C NMR spectrum is recorded in DMSO using TMS as an internal standard shown in (Figures 4.9–4.16). The chemical shift values of different protons obtained are as in (Table 4.4):

COMPOUND	ASSIGNMENTS
QPBA	^1H NMR (DMSO δ ppm); 6.2 (Ar-H), 6.4 (1H, Ar-H), 6.8 (1H, Ar-H), 7.5 (1H, Ar-H), 7.8 (1H, Ar-H), 9.4 (2H, OH), 9.7 (1H, OH), 10.9 (1H, OH), 12.5 (1H, OH). ^{13}C NMR (DMSO δ ppm); 93.84 (Ar-C), 98.62 (Ar-C), 103.46 (Ar-C), 115.49, 116.05 (Ar-C), 120.42 (Ar-C), 122.39 (A2C), 136.19 (ArOH), 145.50 (Ar-OH), 147.23 (Ar-C-OH), 148.15 (Ar-CO), 156.57 (Ar-CO), 161.17 (Ar-C-OH), 164.33 (Ar-C-OH), 176.29 (ArC=N).

QANP	<p>^1H NMR(DMSO-d_6 ppm); 5.2 (1H, Ar-H), 6.2 (1H, Ar-H), 6.4 (1H, Ar-H), 6.8 (1H, Ar-H), 6.9 (1H, Ar-H), 7.45 (1H, Ar-H), 7.7 (1H, Ar-H), 9.4 (2H, Ar-OH), 9.6 (1H, Ar-OH), 10.8 (1H, Ar-OH), 12.5 (1H, Ar-OH).</p> <p>^{13}C NMR(DMSO-d_6 ppm); 93.80 (Ar-C), 98.62 (Ar-C), 103.45 (Ar-C), 108.09 (Ar-C), 113.53 (Ar-C), 113.56 (Ar-C), 115.49 (Ar-C), 116.05 (Ar-C), 120.42 (Ar-C), 122.39 (Ar-C), 136.19 (Ar-C-OH), 138.07 (Ar-C-OH), 140.61 (Ar-C-OH), 145.50 (Ar-C-OH), 147.03 (Ar-C-OH), 148.14 (Ar-C-OH), 151.16 (Ar-C-OH), 156.57 (Ar-C=O), 176.29 (C=N).</p>
QOPD	<p>^1H NMR(DMSO-d_6 ppm); 6.1 (2H, Ar-H), 6.3 (3H, Ar-H), 6.5 (2H, Ar-H), 6.8 (3H, Ar-H), 6.9 (2H, Ar-H), 7.5 (2H, Ar-H), 7.7 (2H, Ar-H), 9.4 (5H, Ar-OH), 12.5 (2H, Ar-OH).</p> <p>^{13}C NMR(DMSO-d_6 ppm); 93.80 (Ar-C), 98.66 (Ar-C), 103.46 (Ar-C), 115.04 (Ar-C), 115.52 (Ar-C), 116.05 (Ar-C), 117.79 (Ar-C), 120.44 (Ar-C), 122.42 (Ar-C), 135.36 (Ar-C), 136.19 (Ar-C-OH), 145.51 (Ar-C-OH), 147.24 (Ar-C-OH), 148.15 (Ar-C-OH), 156.60 (Ar-C-OH), 161.18 (Ar-C=O), 164.37 (Ar-CO), 176.29 (C=N).</p>

QAMP	<p>^1HNMR(DMSO-d_6); 2.07(3H, ArH), 4.45(1H, Ar-H), 6.2 (1H, Ar-H), 6.4 (2H, Ar-H), 6.6(1H, Ar-H), 6.9(1H, Ar-H), 7.6(1H, ArH), 7.7(1H, ArH) 8.67(1H, Ar-H), 9.37(3H, Ar-OH), 12.5(1H, Ar-OH).</p> <p>^{13}CNMR(DMSO-d_6); 93.81(Ar-C), 98.64(Ar-C), 103.45(Ar-C), 114.69(Ar-C), 115.51(Ar-C), 115.68(Ar-C), 116.05(ArC), 117.19(Ar-C), 120.43(Ar-C), 122.41(Ar-C), 136.19(Ar-C-OH), 136.69 (Ar-C-OH), 142.21(Ar-C-OH), 145.51(Ar-C-OH), 147.24(Ar-C-OH), 148.15(Ar-C-OH), 156.59(Ar-C-O), 161.17(Ar-CO), 176.29 (C=N).</p>
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Table 4.4: ^1H and ^{13}C NMR spectral data of the Schiff BasesFigure 4.9: ^1H NMR spectrum of QPBA

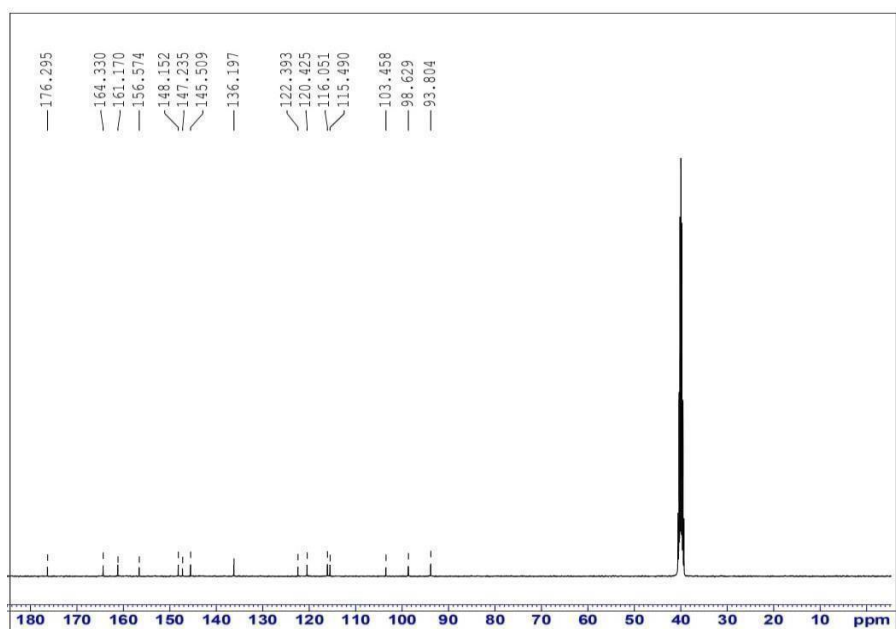


Figure 4.10: ¹³C NMR spectrum of QPBA

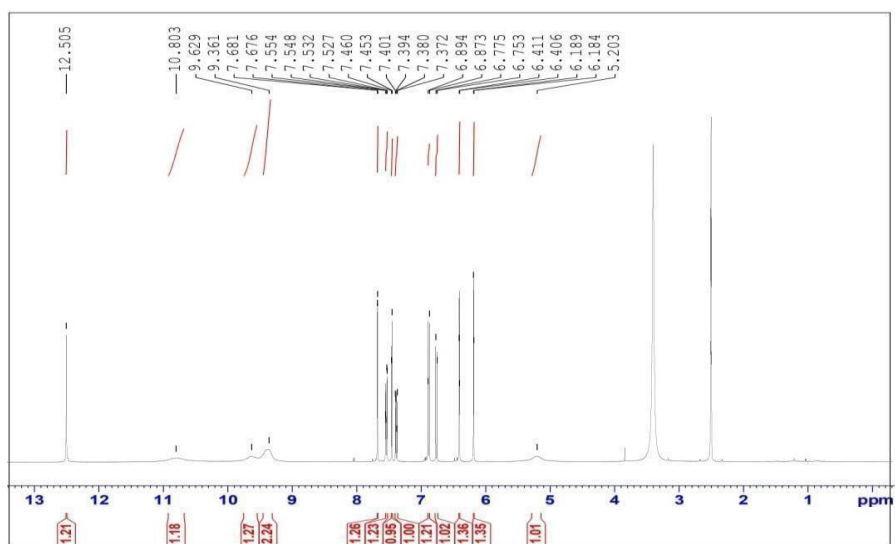


Figure 4.11: ¹H NMR spectrum of QANP

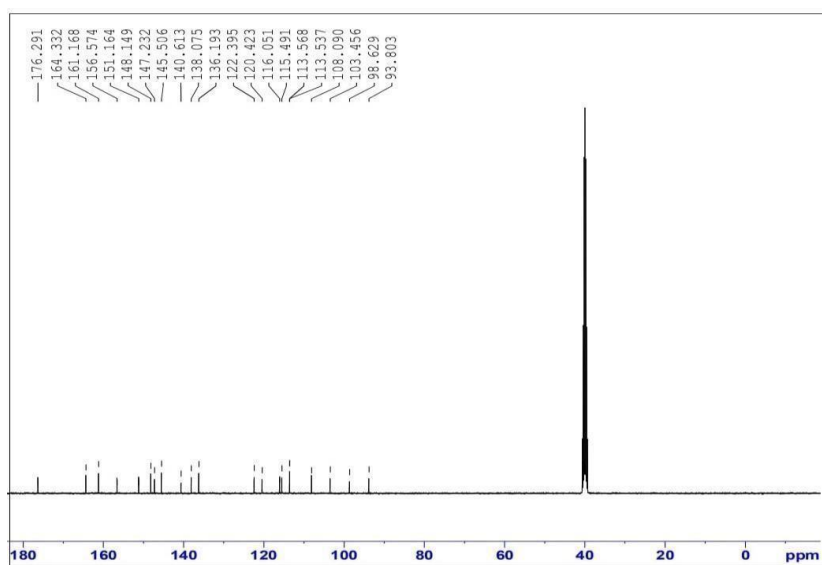


Figure 4.12: ^{13}C NMR spectrum of QANP

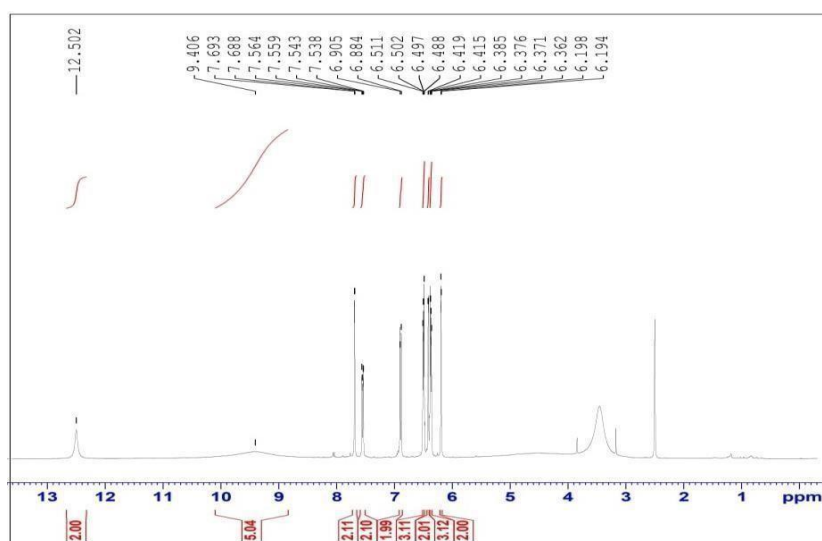


Figure 4.13: ^1H NMR spectrum of QOPD

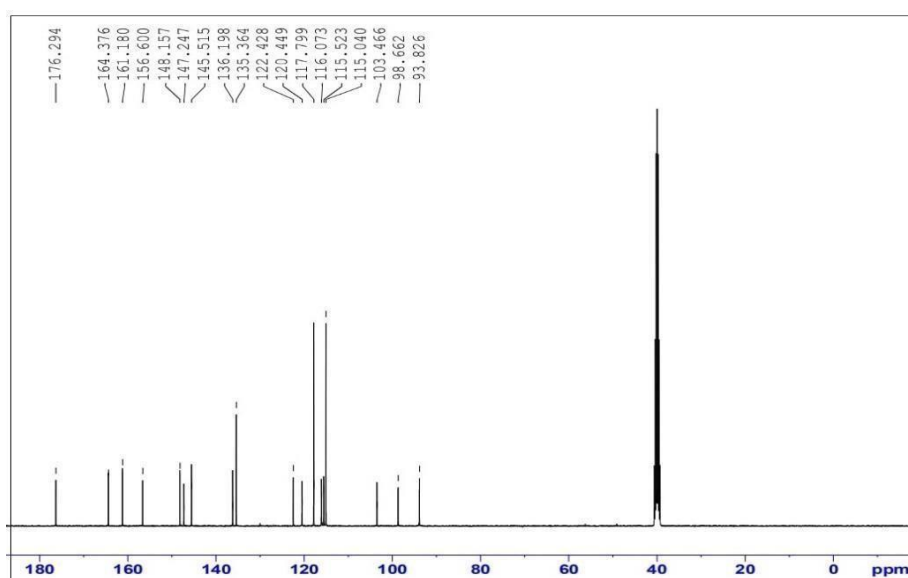


Figure 4.14: ¹³C NMR spectrum of QOPD

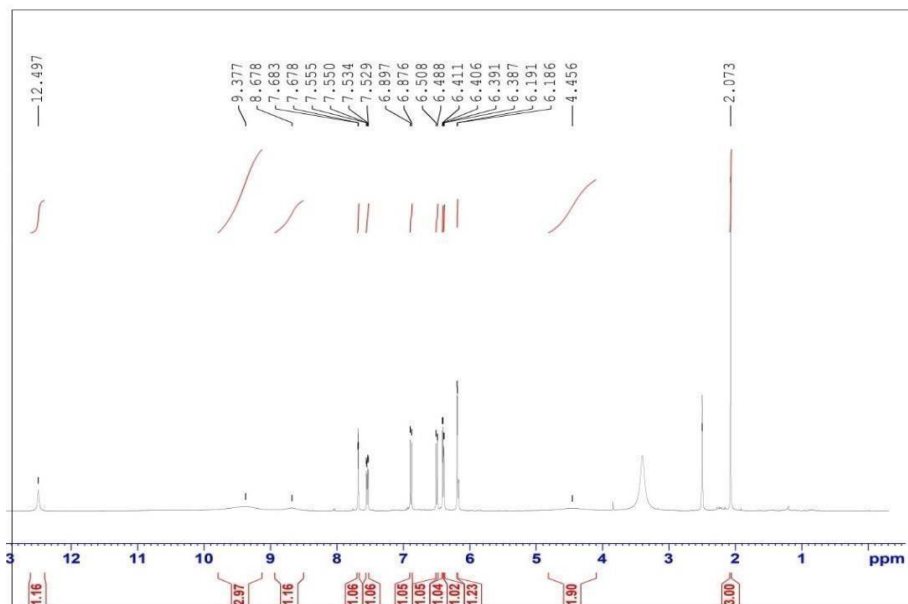


Figure 4.15: ¹H NMR spectrum of QAMP

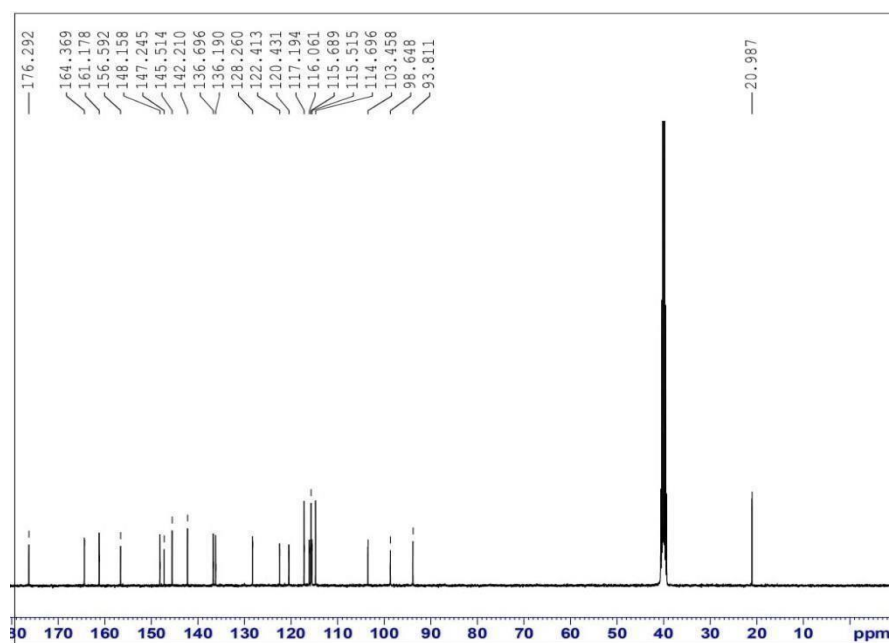


Figure 4.16: ^{13}C NMR spectrum of QAMP

4.1.5 ANTI-INFLAMMATORY ACTIVITY

The anti-inflammatory activity of the Quercetin Schiff Bases was studied using the HRBC membrane stabilization method. The % of inhibition of the four compounds at different concentrations was calculated. The percentage of inhibition values of QPBA, QANP, QOPD, QAMP were tabulated in Tables 4.5-4.8. To find the IC₅₀ value of the Quercetin Schiff bases a graph was plotted with values % of inhibition against the concentration of the sample which is shown in (Figures 4.16-4.18).

QPBA

Concentration of sample (mg/ml)	% of Inhibition
2.5	34.5
5	57
10	74.9

Table 4.5: Anti-inflammatory activity of QPBA

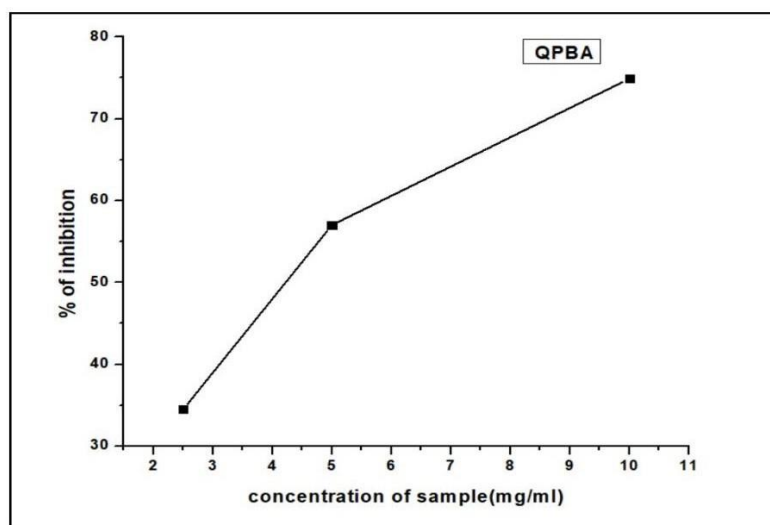


Figure4.16:Anti-inflammatoryofQPBA

QANP

Concentration of sample(mg/ml)	%ofInhibition
2.5	34.7
5	40.4
10	54.9

Table4.6:Anti-inflammatoryactivityofQANP

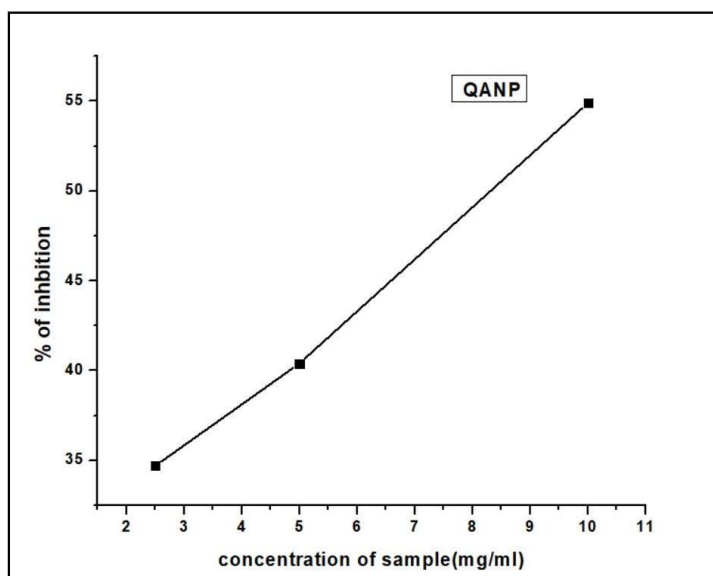


Figure 4.17: Anti-inflammatory activity of QANP QOPD

Concentration of sample(mg/ml)	%ofInhibition
2.5	34.6
5	42.3
10	53.1

Table 4.7: Anti-inflammatory activity of QOPD

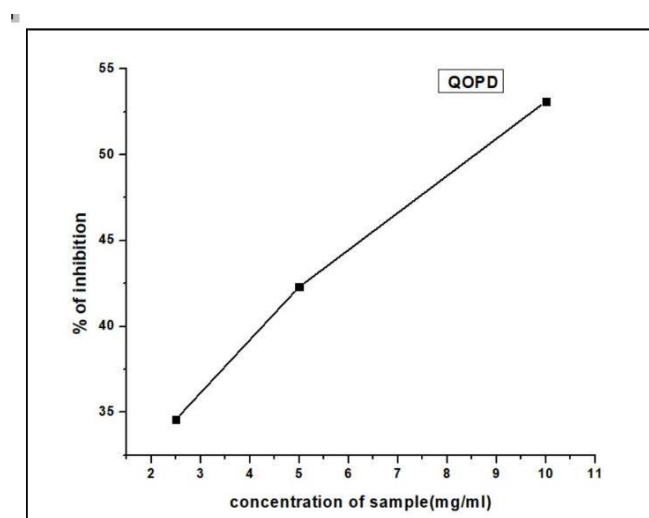


Figure4.17:Anti-inflammatoryofQOPD

QAMP

Concentration of sample(mg/ml)	%ofInhibition
2.5	32.4
5	41
10	51.2

Table4.8:Anti-inflammatoryactivityofQAMP

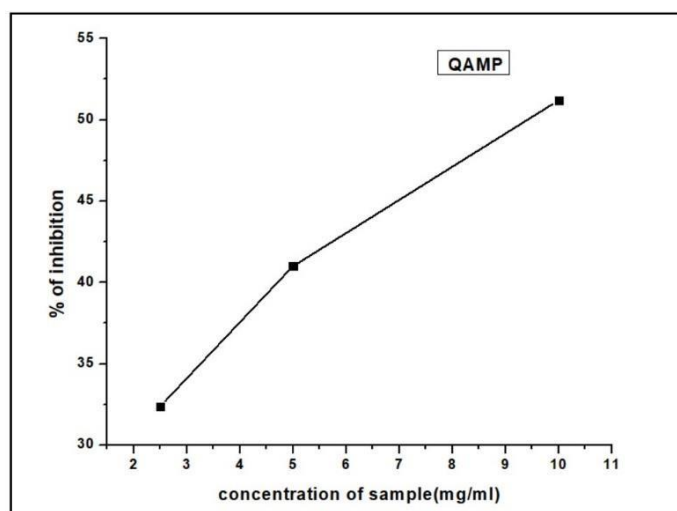


Figure 4.18: Anti-inflammatory effect of QAMP IC_{50}

VALUES OF SCHIFF BASES

IC_{50} represents the concentration at which a substance exerts half of its maximal inhibitory effect. This value is typically used to characterize the effectiveness of an antagonist in inhibiting a specific biological or biochemical process. Low IC_{50} value means that the compound is potent at low concentrations, and thus will show lower systemic toxicity when administered to the patient [50]. The IC_{50} values of QPBA, QANP, QOPD and QAMP are given in the (Table 4.9):

COMPOUND	IC_{50} VALUES (mg/ml)
QPBA	1.4978
QANP	1.5147
QOPD	1.4662
QAMP	1.4810
Ibuprofen	2.6817

TABLE 4.9: IC_{50} VALUES OF SCHIFF BASES

These IC₅₀ values of Schiff bases were compared with ibuprofen a widely used non-steroidal AntiInflammatorydrug. The IC₅₀ value of ibuprofen is 2.6817 mg/ml The IC₅₀ value of all the synthesized Schiff Bases are much lower than that of ibuprofen.

Chapter 5

Conclusions

In the present work four new Quercetin Schiff bases were synthesised and characterized. These are synthesized by the condensation of Parabromo aniline, 2-amino-4-nitrophenol, Ortho phenylenediamine and 2-Amino-4-methylphenol with Quercetin. Synthesized Schiff bases are characterised by IR, UV-visible, ^1H NMR and ^{13}C NMR spectroscopic method. Anti-inflammatory activity of the Schiff bases are evaluated. The IC_{50} values of Schiff bases were compared with ibuprofen, a widely used non-steroidal anti-inflammatory drug. From the study QPBA, QANP, QOPD, QAMP is found to be more effective than ibuprofen and among the four Schiff bases QOPD is more effective for Anti-inflammatory activity because of its lower IC_{50} value.

References

- [1] N.S.A.Xavier, "Synthesis and study of Schiff base ligands," *IOSR J. Appl. Chem.*, 2024.
- [2] N. K. Chaudhary, B. Guragain, S. K. Chaudhary, and P. Mishra, "Schiff base metal complex as a potential therapeutic drug in medical science: A critical review," *Bibechana*, vol. 18, no. 1, pp. 214–230, 2021, doi: 10.3126/bibechana.v18i1.29841.
- [3] A.S. and P.A. Solankee, Thakur, "Synthesis, Antibacterial and Antifungal Activities of s-Triazine Derivatives," *E-Journal Chem.*.
- [4] X. Ren, K. Liu, Y. Lu, W. Ding, S. Lei, and L. Yang, "Stepwise versus Concerted: Theoretical Insights into the Stereoselectivity in Aryl Imine Formation Assisted by Acid and Water," *J. Phys. Chem. A*, vol. 127, no. 46, pp. 9748–9759, 2023, doi: 10.1021/acs.jpca.3c05530.
- [5] A. Soroceanu and A. Bargin, "Advanced and Biomedical Applications of Schiff-Base Ligands and Their Metal Complexes: A Review," *Crystals*, vol. 12, no. 10, 2022, doi: 10.3390/cryst12101436.
- [6] D. A. Xavier and N. Srividhya, "Synthesis and Study of Schiff base Ligands," *IOSR J. Appl. Chem.*, vol. 7, no. 11, pp. 06–15, 2014, doi: 10.9790/5736-071110615.

- [7] A. Altameemi, K. K. Abid, R. H. Al -Bayati, and A. A. Faeq, "Transition Metal Complexes of New N-AminoQuinolone Derivative; Synthesis, Characterization, Thermal Study and Antimicrobial Properties," *Am. J. Chem.*, vol. 6, no. 2, pp. 29–35, 2016, [Online]. Available: <http://journal.sapub.org/chemistry>
- [8] T. Maharana, N. Nath, H. C. Pradhan, S. Mantri, A. Routaray, and A. K. Sutar, "Polymer-supported first-row transition metal schiff base complexes: Efficient catalysts for epoxidation of alkenes," *React. Funct. Polym.*, vol. 171, 2022, doi: 10.1016/j.reactfunctpolym.2021.105142.
- [9] A. Alsayari *et al.*, "Synthesis, characterization, and biological evaluation of some novel pyrazolo[5,1-b]thiazole derivatives as potential antimicrobial and anticancer agents," *Molecules*, vol. 26, no. 17, 2021, doi: 10.3390/molecules26175383.
- [10] Z. Guo *et al.*, "The synthesis and antioxidant activity of the Schiff bases of chitosan and carboxymethyl chitosan," *Bioorganic Med. Chem. Lett.*, vol. 15, no. 20, pp. 4600–4603, 2005, doi: 10.1016/j.bmcl.2005.06.095.
- [11] H. Guo, J. B. Callaway, and J. P. Y. Ting, "Inflammasomes: Mechanism of action, role in disease, and therapeutics," *Nat. Med.*, vol. 21, no. 7, pp. 677–687, 2015, doi: 10.1038/nm.3893.
- [12] X. Zhang *et al.*, "Mitochondrial DNA in liver inflammation and oxidative stress," *Life Sci.*, vol. 236, 2019, doi: 10.1016/j.lfs.2019.05.020.
- [13] S. B. Ade, D. G. Kolhatkar, and M. N. Deshpande, "Synthesis, characterization and biological activity of a schiff base derived

- fromisatin and 2-amino, 4-methyl phenol and its transition metal complexes,” *Int. J. Pharma Bio Sci.*, vol. 3, no. 2, pp. 350–356, 2012.
- [14] K. Venkatesh, L. Venkata Reddy, K. B. Chandra Sekhar, and K. Mukkanti, “Synthesis, characterization and antimicrobial activity of transition metal complexes of schiff base,” *Int. J. ChemTech Res.*, vol. 3, no. 2, pp. 676–679, 2011.
- [15] T. Daniel Thangadurai, M. Gowri, and K. Natarajan, “Synthesis and characterisation of ruthenium(III) complexes containing monobasic bidentate Schiff bases and their biological activities,” *Synth. React. Inorg. Met. Chem.*, vol. 32, no. 2, pp. 329–343, 2002, doi: 10.1081/SIM-120003211.
- [16] P. M. Kearney, C. Baigent, J. Godwin, H. Halls, J. R. Emberson, and C. Patrono, “Do selective cyclo-oxygenase-2 inhibitors and traditional non-steroidal anti-inflammatory drugs increase the risk of atherothrombosis? Meta-analysis of randomised trials,” *Br. Med. J.*, vol. 332, no. 7553, pp. 1302–1305, 2006, doi: 10.1136/bmj.332.7553.1302.
- [17] G. W. Hanks, “Gastrointestinal toxicity of different nonsteroidal anti-inflammatory drugs”, *Journal of Pain and Symptom Management*”.
- [18] M. S. Alam and J. U. Ahmed, “Synthesis, crystal structure, biological evaluation, in silico ADME properties, enzymatic target prediction and molecular docking studies of pyrazolone-azomethine analogs,” *J. Mol. Struct.*, vol. 1294, 2023, doi: 10.1016/j.molstruc.2023.136504.

References

- [19] V.Lobo,A.Patil,A.Phatak,andN.Chandra,“Freeradicals, antioxidantsandfunctionalfoods:Impactonhumanhealth,” *Pharmacogn.Rev.*,vol.4,no.8,pp.118–126,2010,doi: 10.4103/0973-7847.70902.
- [20] L.I.Soce, D. C. Visan, S. F. Barbuceanu, T. V. Apostol, O. G. Bratu, and B. Socea, “The antioxidant activity of some acylhydrazones with dibenzo [a,d][7] annulene moiety,” *Rev.Chim.*, vol. 69, no. 4, pp. 795–797, 2018, doi: 10.37358/rc.18.4.6202.
- [21] M. M. Rahman, M. B. Islam, M. Biswas, and A. H. M. Khurshid Alam, “In vitro antioxidant and free radical scavenging activity of different parts of *Tabebuia pallida* growing in Bangladesh,” *BMC Res. Notes*, vol. 8, no. 1, 2015, doi: 10.1186/s13104-015-1618-6.
- [22] E. B. Kurutas, “The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: Current state,”*Nutr.J.*,vol.15,no.1,2016,doi:10.1186/s12937-016-0186- 5.
- [23] G. A. Engwa, “Free Radicals and the Role ofPlant Phytochemicals as Antioxidants Against Oxidative Stress-RelatedDiseases,” *Phytochem. - Source Antioxidants Role Dis. Prev.*, 2018, doi: 10.5772/intechopen.76719.
- [24] M. Taghvaei and S. M. Jafari, “Application and stability of natural antioxidantsinedible oils inorderto substitute synthetic additives,” *J. Food Sci. Technol.*, vol. 52, no. 3, pp. 1272–1282, 2015, doi: 10.1007/s13197-013-1080-1.
- [25] R.F.CarlosA.M.Afonso,NunoR.Candeias,DulcePereira

- Simão, Alexandre F. Trindade, Jaime A. S. Coelho, Bin Tan, "Comprehensive Organic Chemistry Experiments for the Laboratory Classroom," *Compr. Org. Chem. Exp. Lab. Classr.*, 2016, doi: 10.1039/9781839168673.
- [26] L. Mukhtarov, G. Pestsov, M. Nikishina, E. Ivanova, Y. Atroshchenko, and L. Perelomov, "Fungicidal Properties of 2-Amino-4-nitrophenol and Its Derivatives," *Bull. Environ. Contam. Toxicol.*, vol. 102, no. 6, pp. 880–886, 2019, doi: 10.1007/s00128-019-02602-4.
- [27] M. Vedamalai *et al.*, "Carbon nanodots prepared from o-phenylenediamine for sensing of Cu²⁺ ions in cells," *Nanoscale*, vol. 6, no. 21, pp. 13119–13125, 2014, doi: 10.1039/c4nr03213f.
- [28] C. Fischer, V. Speth, S. Fleig-Eberenz, and G. Neuhaus, "Induction of zygotic polyembryos in wheat: Influence of auxin polar transport," *Plant Cell*, vol. 9, no. 10, pp. 1767–1780, 1997, doi: 10.1105/tpc.9.10.1767.
- [29] B.M. Mohammed *et al.*, "Vitamin C promotes wound healing through novel pleiotropic mechanisms," *Int. Wound J.*, vol. 13, no. 4, pp. 572–584, 2016, doi: 10.1111/iwj.12484.
- [30] G.F. DEGAETANI, "Chronic inflammation," *Rass. Clin. Sci.*, vol. 39, pp. 43–48, 1963, doi: 10.5005/jp/books/12977_5.
- [31] M. Mokhtari, R. Razzaghi, and M. Momen-Heravi, "The effects of curcumin intake on wound healing and metabolic status in patients with diabetic foot ulcer: A randomized, double-blind, placebo-controlled trial," *Phyther. Res.*, vol. 35, no. 4, pp. 2099–2107, 2021, doi: 10.1002/ptr.6957.

References

- [32] R.J.Flower, “Studies on the mechanism of action of anti-inflammatory drugs: A paper in honour of John Vane,” *Thromb. Res.*, vol. 110, no. 5–6, pp. 259–263, 2003, doi: 10.1016/S0049-3848(03)00410-9.
- [33] A. Sodhi, S. Naik, A. Pai, and A. Anuradha, “Rheumatoid arthritis affecting temporomandibular joint,” *Contemp. Clin. Dent.*, vol. 6, no. 1, pp. 124–127, 2015, doi: 10.4103/0976-237X.149308.
- [34] C. Tian *et al.*, “Investigation of the anti-inflammatory and antioxidant activities of luteolin, kaempferol, apigenin and quercetin,” *South African J. Bot.*, vol. 137, pp. 257–264, 2021, doi: 10.1016/j.sajb.2020.10.022.
- [35] M. Lesjak *et al.*, “Antioxidant and anti-inflammatory activities of quercetin and its derivatives,” *J. Funct. Foods*, vol. 40, pp. 68–75, 2018, doi: 10.1016/j.jff.2017.10.047.
- [36] A. P. Rogerio *et al.*, “Anti-inflammatory activity of quercetin and isoquercitrin in experimental murine allergic asthma,” *Inflamm. Res.*, vol. 56, no. 10, pp. 402–408, 2007, doi: 10.1007/s00011-007-7005-6.
- [37] Y. Sakanashi *et al.*, “Possible use of quercetin, an antioxidant, for protection of cells suffering from overload of intracellular Ca^{2+} : A model experiment,” *Life Sci.*, vol. 83, no. 5–6, pp. 164–169, 2008, doi: 10.1016/j.lfs.2008.05.009.
- [38] S. A. N. Aljadaan, R. S. Elias, and R. A. Al-Anssari, “Investigation of the antioxidant and antibacterial activity of novel quercetin derivatives,” *Biointerface Res. Appl. Chem.*, vol. 10, no. 6, pp. 7329–7336, 2020, doi: 10.33263/BRIAC106.73297336.

- [39] A. Dehghani, A. H. Mostafatabar, G. Bahlakeh, B. Ramezanzadeh, and M. Ramezanzadeh, "Detailed-level computer modeling explorations complemented with comprehensive experimental studies of Quercetin as a highly effective inhibitor for acid-induced steel corrosion," *J. Mol. Liq.*, vol. 309, 2020, doi: 10.1016/j.molliq.2020.113035.
- [40] A. R. Mahmud *et al.*, "Natural flavonols: actions, mechanisms, and potential therapeutic utility for various diseases," *Beni-Suef Univ. J. Basic Appl. Sci.*, vol. 12, no. 1, 2023, doi: 10.1186/s43088-023-00387-4.
- [41] M.S. Refat *et al.*, "Potential therapeutic effects of new ruthenium (III) complex with quercetin: Characterization, structure, gene regulation, and antitumor and anti-inflammatory studies (RuIII/Q novel complex is a potent immunoprotective agent)," *Crystals*, vol. 11, no. 4, 2021, doi: 10.3390/cryst11040367.
- [42] E. Frąckowiak, A. Płatek-Mielczarek, J. Piwek, and K. Fic, "Advanced characterization techniques for electrochemical capacitors," *Adv. Inorg. Chem.*, vol. 79, pp. 147–203, 2022, doi: 10.1016/bs.adioch.2021.12.006.
- [43] B. H. Stuart, "Infrared Spectroscopy: Fundamentals and Applications."
- [44] B. M. Weckhuysen, "In-situ spectroscopy of catalysts."
- [45] A. Raja, Pavan; Barron, "Physical Methods in Chemistry and Nano Science," *OpenStax CNX*, 2019, [Online]. Available: [https://espanol.libretexts.org/Quimica/Química_Analítica/Métodos_Físicos_en_Química_y_Nano_Ciencia_\(Barron\)/01%3A_Análisis_](https://espanol.libretexts.org/Quimica/Química_Analítica/Métodos_Físicos_en_Química_y_Nano_Ciencia_(Barron)/01%3A_Análisis_)

- Elemental/1.04%3A_Introducción_a_la_Espectroscopia_de_Absorción_Atómica
- [46] F. REXHEPI, G. KURTI, F. FERATI, and S. SHALA, “Ftir-Spectroscopy Study of Microwave and Conventional Heating on the Degradation of Margarine and Butter,” *Eur. J. Mater. Sci. Eng.*, vol. 4, no. 1, pp. 3–10, 2019, doi: 10.36868/ejmse.2019.04.01.003.
- [47] A. Synytsya, J. Čopíková, P. Matějka, and V. Machovič, “Fourier transform Raman and infrared spectroscopy of spectins,” *Carbohydr. Polym.*, vol. 54, no. 1, pp. 97–106, 2003, doi: 10.1016/S0144-8617(03)00158-9.
- [48] L. Duan *et al.*, “A novel and versatile precursor for the synthesis of highly preorganized tetradentate ligands based on phenanthroline and their binding properties towards lanthanides(III) ions,” *Colloids Surfaces A Physicochem. Eng. Asp.*, vol. 647, 2022, doi: 10.1016/j.colsurfa.2022.129089.
- [49] *etal.*, “Ftir-Spectroscopy Study of Microwave and Conventional Heating on the Degradation of Margarine and Butter,” *Eur. J. Mater. Sci. Eng.*, vol. 4, no. 1, pp. 3–10, 2019, doi: 10.36868/ejmse.2019.04.01.003.
- [50] S. Rajan and S. S. Sidhu, “Simplified synthetic antibody libraries,” *Methods Enzymol.*, vol. 502, pp. 3–23, 2012, doi: 10.1016/B978-0-12-416039-2.00001-X.